

# Analyzing cannabis flowers according to the German Pharmacopeia - monograph 2017

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## SUMMARY

Since the change in German law in 2017 (§1 Abs. 1, BtMG) [1] the need for reliable and robust HPLC methods for quality control has drastically increased. Six common cannabinoids of high medicinal interest cannabidiol (CBD), cannabidiolic acid (CBDA), cannabinol (CBN),  $\Delta^8$ -tetrahydrocannabinol ( $\Delta^8$ -THC),  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC), and  $\Delta^9$ -tetrahydrocannabinolic acid ( $\Delta^9$ -THCA) were quantified using the AZURA® HPLC plus system according to the monography of German Pharmacopeia [2]. Three different reversed phase columns were screened for robustness for the method verified in the subsequent step. Method verification took place using authorized medicinal cannabis flower samples available on the market. The substances were analysed with the help of chemical reference standards.

## INTRODUCTION

*Cannabis sativa* L. is one of the oldest agricultures and medicinal plant which produces a variety of compounds such as terpenoids, flavonoids and cannabinoids [3]. The interaction of cannabinoids with the body's own cannabinoid receptors, which occur in a variety of brain cells for coordination, memory processing and spatial orientation, opens up new pharmacological and psychological treatment options [4]. Probably the most psychoactive cannabinoid of the four different isomers of  $\Delta^9$ -THC is the (-)- $\Delta^9$ -trans-tetrahydrocannabinol, also known as dronabinol. In Germany, due to its psychoactive properties,  $\Delta^9$ -THC is controlled by the narcotics law (from the German Betäubungsmittelschutzgesetz (BtMG)). In

March 2017, the regulations changed by the amendment of article 1 BtMG. The amendment of annexes II and III of the

BtMG now allows cannabis such as marijuana plants and plant parts to be marketed and prescribed and are therefore authorised for medical purposes as ready-to-use medicinal products [1]. As cannabis is an approved medicine, production must be conducted and monitored in accordance with Good Manufacturing Practice (GMP) guidelines. In order to confirm this quality and to guarantee accurate labelling of medicinal products, food and cosmetics, the demand for standardized methods for the quantitative and qualitative determination of the ingredients especially the cannabinoids is increasing [6]. The quality assurance of the plants may be ensured by employing the German Pharmacopeia method DAB [1,5]. In this work, the HPLC method for cannabis flowers according to the DAB monography including robustness evaluation was carried out with the AZURA HPLC plus system.



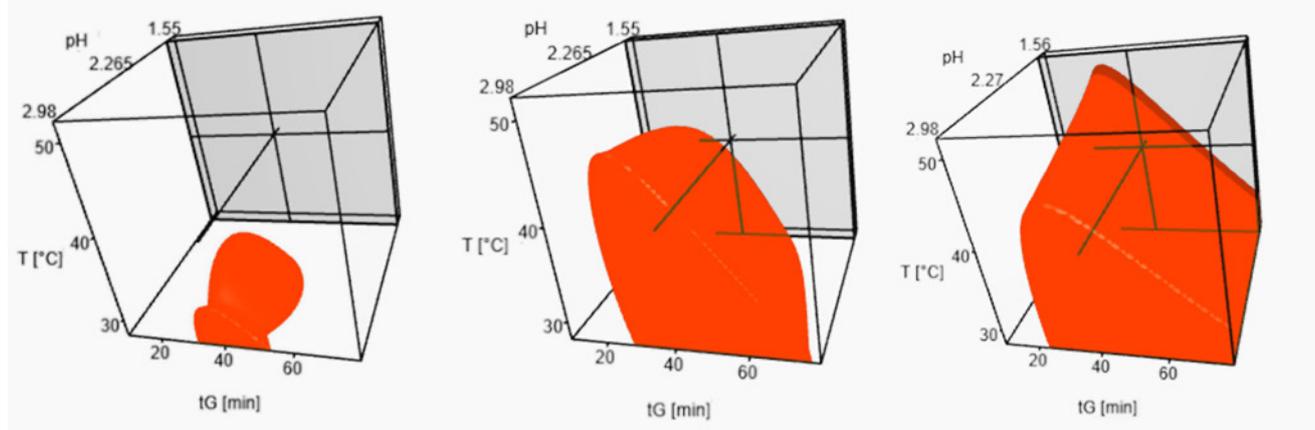
Additional Information

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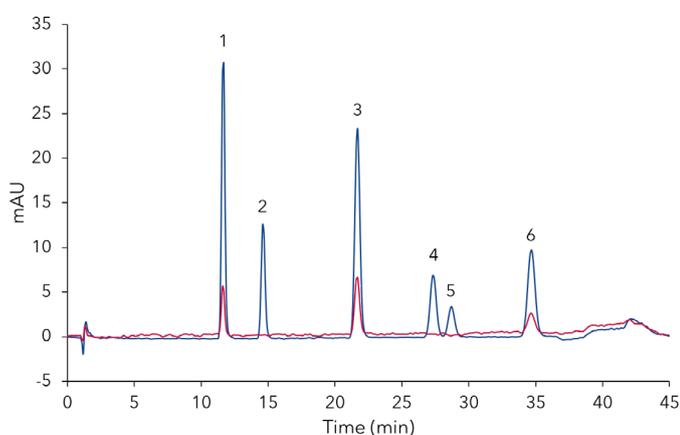
## RESULTS

A combined mobile and stationary phase design of experiment (DoE) approach with the aid of the HPLC modelling software DryLab (Molnár-Institute, Berlin, Germany) was employed. Investigated parameters were gradient time (tG), temperature (T), pH and the stationary phase of the column, producing the three dimensional spaces shown in Fig 1. The red areas in Fig 1 display the robust space of each column (critical resolution > 1.6), from which it is easily seen that the largest area is represented by the column C18P on the right. Due to the completely filled pH dimension,

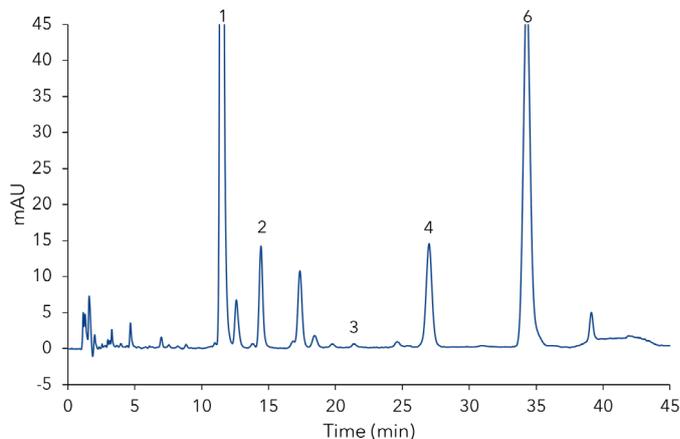
within a given temperature and gradient time, the column C18P shows a nearly unaffected pH stability for this method. Therefore, the column C18P was determined to be most robust and was therefore used for the further measurements. For the assignment of the analytes the resulting chromatogram of a standard mix from the six different cannabinoids with the concentration of 10 µg/mL is shown in Fig 2. The measurement of the cannabis flower bediol (Bedrocan, Veendam, Netherlands) with a dilution of 1 to 10 with ethanol is shown in Fig 3.



**Fig. 1** Representation of the robust space using the tG-T-pH (30/90 min, 30/50°C, pH 1.6/2.2/2.8) with the columns C18 (left), C18H (center) and C18P (right), adjusted robust space level 1.6



**Fig. 2** Chromatogram of standard mix 10 µg/mL, R=1.5 between  $\Delta^8$ -THC and  $\Delta^2$ -THC; blue - 225 nm, red - 306 nm, 1 - CBDA, 2 - CBD, 3 - CBN, 4 -  $\Delta^2$ -THC, 5 -  $\Delta^8$ -THC, 6 -  $\Delta^9$ -THCA



**Fig. 3** Chromatogram of bediol sample 1 mg/mL in ethanol; 1 - CBDA, 2 - CBD, 3 - CBN, 4 -  $\Delta^2$ -THC, 5 -  $\Delta^2$ -THCA

## SAMPLE PREPARATION

The whole 5 g portion of bediol was grinded with a flower grinder. The difference between grinded and original bediol is shown in Figure 5. The sample preparation was performed according to DAB, where 500 mg substance was extracted three times with 15 mL ethanol on a laboratory shaker for 15 min with a following centrifugation at 5000 rpm for one minute. Mixing all the extracts within 50 mL measuring flask, a 1:10 dilution was carried out and measured with the HPLC system after filtrating over a 0.45  $\mu\text{m}$  RC filter.



Fig.4 Bediol sample (left original, right grinded)

## CONCLUSION

A design of experiment combined with a column screening can be a helpful tool to determine the robustness of a method. With the help of the HPLC Modelling Software Drylab this could be tested within the three different columns with convincing results. The measured chromatograms show a sufficient separation of the six given cannabinoids according to DAB method. The specification of the DAB with a critical resolution of  $R > 1.2$  for the critical analyte pair  $\Delta^8$ -THC and  $\Delta^9$ -THC is confirmed with  $R = 1.5$  within the measurements. The sample measurement of bediol shows a sufficient assignment of the signals whereas the not identified signals can be assumed as matrix or not categorized cannabinoids. Additionally, the use of two different wavelengths shows a differentiation between the acid and neutral forms of the given cannabinoids. CBN also shows an absorption towards 306 nm due to the increased amount of conjugated systems compared to  $\Delta^8$ -THC and  $\Delta^9$ -THC.

## REFERENCES

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## MATERIALS AND METHODS

**Tab. 1** Standard solutions, structures and samples

Analyte	CAS	Purity/ concentration	Manufacturer
Cannabidiol (CBD)	13956-29-1	1.0 mg/mL in MeOH	Cerilliant
Cannabidiolic acid (CBDA)	1244-58-2	99.1% (HPLC)	Sigma- Aldrich
Cannabinol (CBN)	521-35-7	1.0 mg/mL in MeOH	Cerilliant
$\Delta^8$ -Tetrahydrocannabinol ( $\Delta^8$ -THC)	5957-75-5	1.0 mg/mL in MeOH	Cerilliant
$\Delta^9$ -Tetrahydrocannabinol ( $\Delta^9$ -THC)	1972-08-3	1.001 mg/mL in MeOH	Cerilliant
$\Delta^9$ -Tetrahydrocannabinolic acid ( $\Delta^9$ -THCA)	23978-85-0	98.8% (HPLC)	Sigma- Aldrich
Acetonitril	75-05-8	Gradient grade	Merck Millipore
Ethanol	64-17-5	Gradient grade	Merck Millipore
H <sub>3</sub> PO <sub>4</sub>	7664-38-2	AnalaR 85 %	VWR Chemicals

**Tab. 4** System configuration

Instrument	Description	Article no.
Pump	AZURA P 6.1L, LPG	<a href="#">APH34EA</a>
Detector	AZURA MWD 2.1L	<a href="#">ADB01</a>
Flow Cell	10 mm, 10 $\mu$ L, Pressure Proof	<a href="#">AMC38</a>
Autosampler	AZURA AS 6.1L	<a href="#">AAA00AA</a>
Column thermostat	CT 2.1	<a href="#">A05852</a>
Column	Eurospher II 100-3 C18 P, Säule 150 x 4.6 mm	<a href="#">15VE182E2G</a>
	Eurospher II 100-3 C18 H, Säule 150 x 4.6 mm	<a href="#">15VE185E2G</a>
	Eurospher II 100-3 C18, Säule 150 x 4.6 mm	<a href="#">15VE181E2G</a>
Software	Clarity Chrom 8.1	<a href="#">A1670</a>
HPLC modelling software	DryLab 4.3.4.2	-

**Tab. 2** Method parameters

Column temperature	40°C
Injection volumen	10 $\mu$ L
Injection mode	Full loop
Detection	UV 225 nm /306 nm
Data rate	10 Hz

**Tab. 3** Pump parameters

Eluent A	Water, HPLC grade (H <sub>3</sub> PO <sub>4</sub> 85% 8.64 g/L)		
Eluent B	Acetonitrile, gradient grade		
Flow rate	1.0 mL/min		
Pump program	Time (min)	% A	% B
	0	35	65
	35	30	70
	37	20	80
	40	20	80
	42	35	65
45	35	65	

