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SEMI PREPARATIVE XYLITOL PURIFICATION WITH DEDICATED SUGAR PURIFICATION SYSTEM

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SUMMARY

Xylitol is used as sweetener in the food industry and is generated by chemical conversion of xylose. Here, xylitol was purified from fermentation mash by microbial xylose conversion. The AZURA® sugar purification system with the AZURA RID 2.1L refractive index detector was used for this semi - preparative purification in combination with polymer-based Eurokat Ca column.

INTRODUCTION

The second generation of bio refinery is using biomass with low contents of C6 sugars such as wheat straw. This biomass is often rich in the C5 sugar xylose which is normally not used as a carbon source by microorganisms for ethanol production. Xylose is chemically converted to xylitol which is a five-carbon sugar alcohol occurring in nature mostly in low concentrations and its extraction is too unproductive. It has found its application i.e. food industry as an artificial sweetener in chewing gums. It has been shown that xylose can be converted to xylitol by different yeast and bacteria species [1, 2]. The microbial conversion of xylose to xylitol, followed by a simple purification process, presents an economical and environmentally-friend-ly alternative [3]. A previous study already revealed the feasibility of semi-preparative xylitol purification from fermentation mash (VFD0150). In this study, method optimization for xylitol purification was performed with the same stationary phase material. The AZURA RID 2.1L detector could be used for this task due to its ability to sustain flow rates up to 10 mL/min and 5 bar back pressure.

RESULTS

The separation profile of the semi-preparative Eurokat Ca 150 × 20 mm column was tested by injection of 0.5 mL fermentation mash (FM; 1:2 dilution). Overlay of the resulting chromatogram with chromatograms of standard solution and retention time comparison identified xylitol, mannitol, glycerol and xylose in the sample (see add. results **Fig. A1**). Also at larger injection volumes (1 mL, 2 mL) xylitol could still be baseline separated from mannitol **Fig. 1**. Due to the shorter column length (150 × 20 mm) and faster flow rate (4 mL/min) the xylitol peak

eluted earlier (approx. 13 min) compared to previous study where it eluted at 19 min using a longer column (250 × 16 mm) and lower flow rate (2.5 mL/min) (VFD0150). After injection of 2 mL FM a 12 mL fraction of xylitol was recovered (**Fig. 1**, blue bracket). The analysis of the 12 mL xylitol fraction and subsequent comparison with chromatograms of a xylitol standard (1 mg/mL) and FM revealed no contaminations in the xylitol fraction (**Fig. 2**, red line). Measurements of xylitol concentration in the FM showed an initial concentration of approx. 60 mg/mL xylitol and a concentration of approx. 5.6 mg /mL xylitol in the fraction, revealing an about 11 fold dilution of xylitol by batch purification.

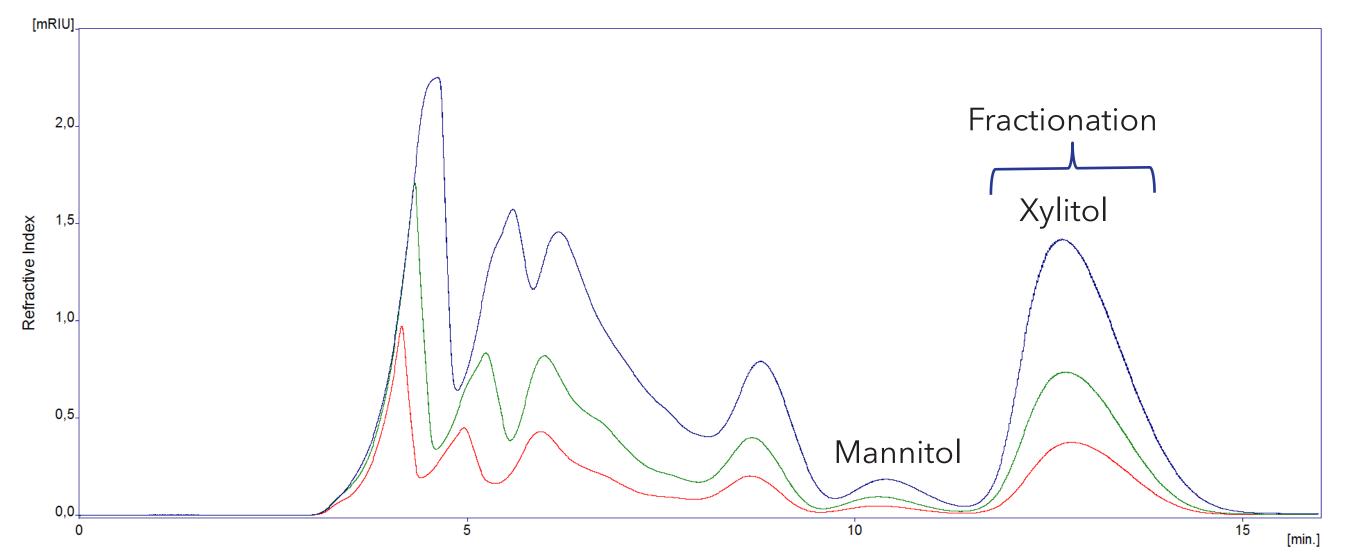


Fig. 1 Chromatogram overlay of different injection volumes from fermentation mash (1:2 dilution); red = 0.5 mL, green - 1 mL, blue = 2 mL; blue brackets-fractionation area 2 mL injection; EK Ca 150 x 20 mm; 4 mL/min; 60 °C

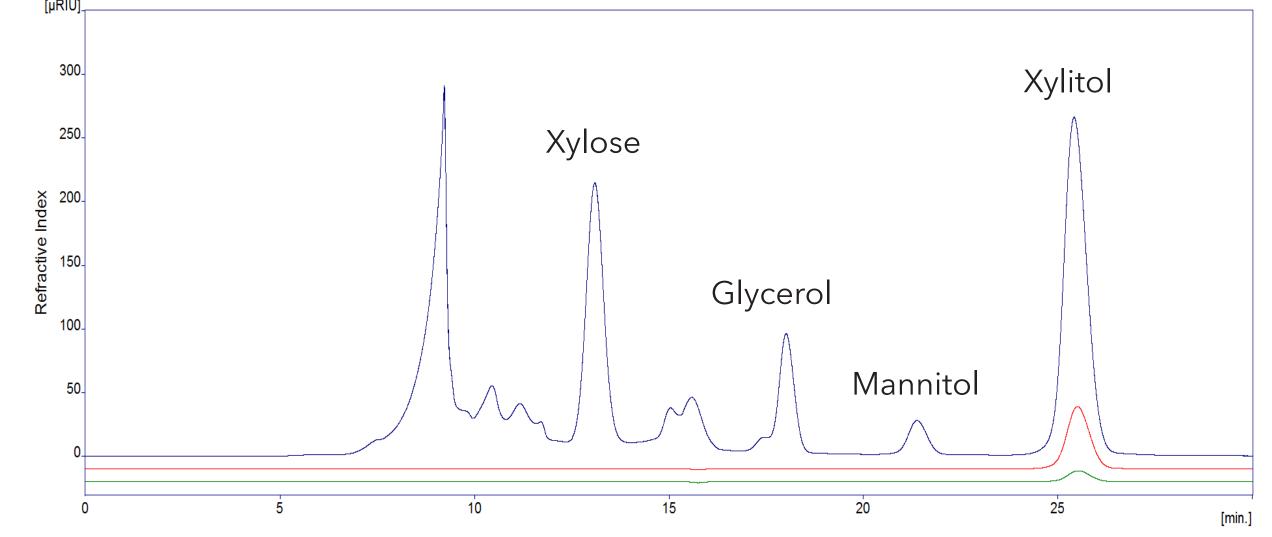


Fig. 2 Overlay of analytical chromatograms; blue - fermantation mash (1:2 dilution); red - fractionation sample from Fig. 1; green - xylitol standard 1 mg/mL; 10 μl each; EK Ca 300 x 8 mm; 75 °C

MATERIALS AND METHOD

The AZURA sugar purification system consists of an assistant AZURA ASM 2.1L with a 12 port multi position valve (for fractionation) and 50 mL pump and an AZURA RID 2.1L refractive index detector. Eurokat Ca 150 × 20 mm column (sulfonated cross-linked styrene-divinylbenzene copolymer) with 25–56 µm particles was used for purification. The column was heated with a heating jacket to 60 °C. Purification run was in isocratic mode for 16 min at 4 mL/min. Different injection volumes were tested. The data rate was set to 5 Hz, time constant 0.02 sec.

CONCLUSION

Two main results were achieved with this study: 1. Optimization of the batch xylitol purification process and 2. Application of the AZURA RID 2.1L refractive index detector for semi-pre-

parative sugar purification at higher flow rates. Xylitol was purified with a purity of >99 % and recovery of >99 % from fermentation mash of microbial xylose to xylitol conversion. Elution time (13 min) and temperature (60 °C) was reduced and injection volume (2 mL) increased when compared to early study (VFD0150).

REFERENCES

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 [3] Chen X., Jian Z.-H., Chen S., Qin W. Int. J. Biol. Sci. 6(7): 834–844 (2010)



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Additional information:

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ADDITIONAL RESULTS

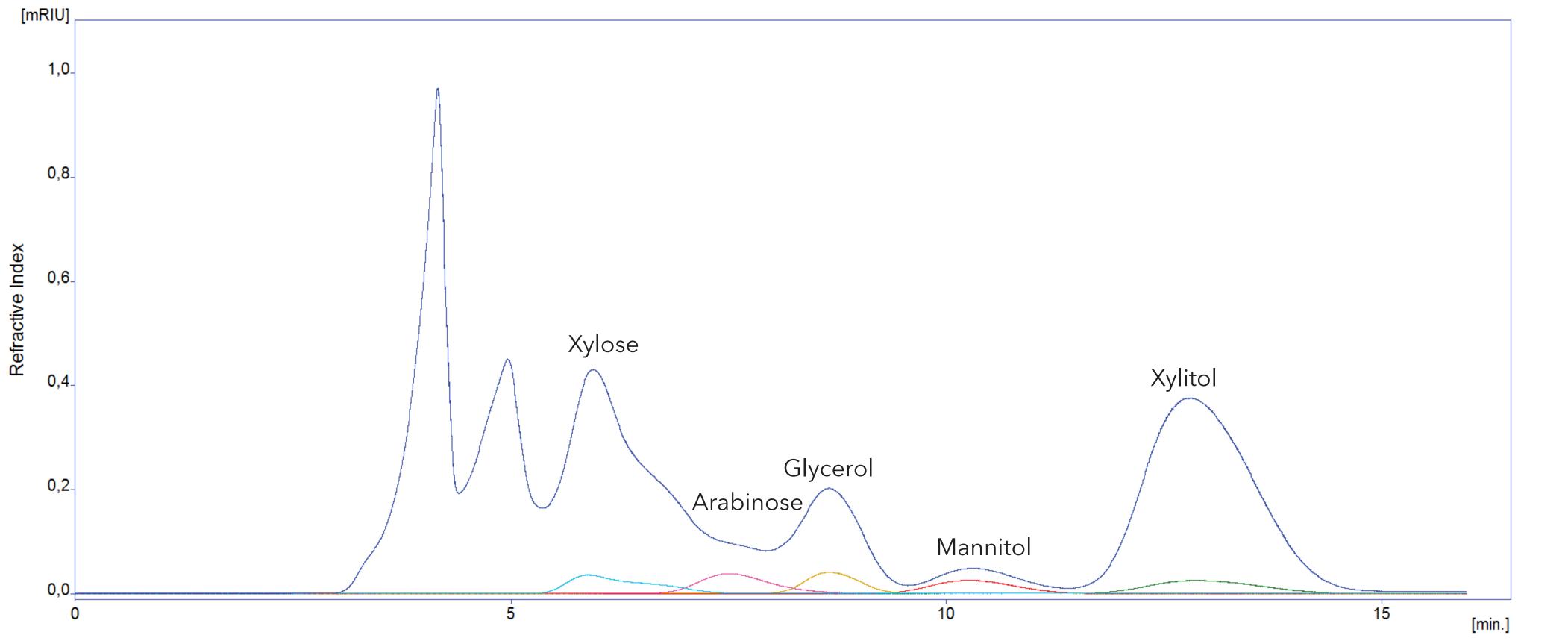


Fig. A1 Chromatograms of 0.5 mL injection of fermentation mash (1:2 dilution) and standards (2 mg/mL each); blue - FM; EK Ca 150 x 20 mm; 60 °C; 4 mL/min

ADDITIONAL MATERIALS AND METHODS

Tab. A1 Method parameters (preparative purification)

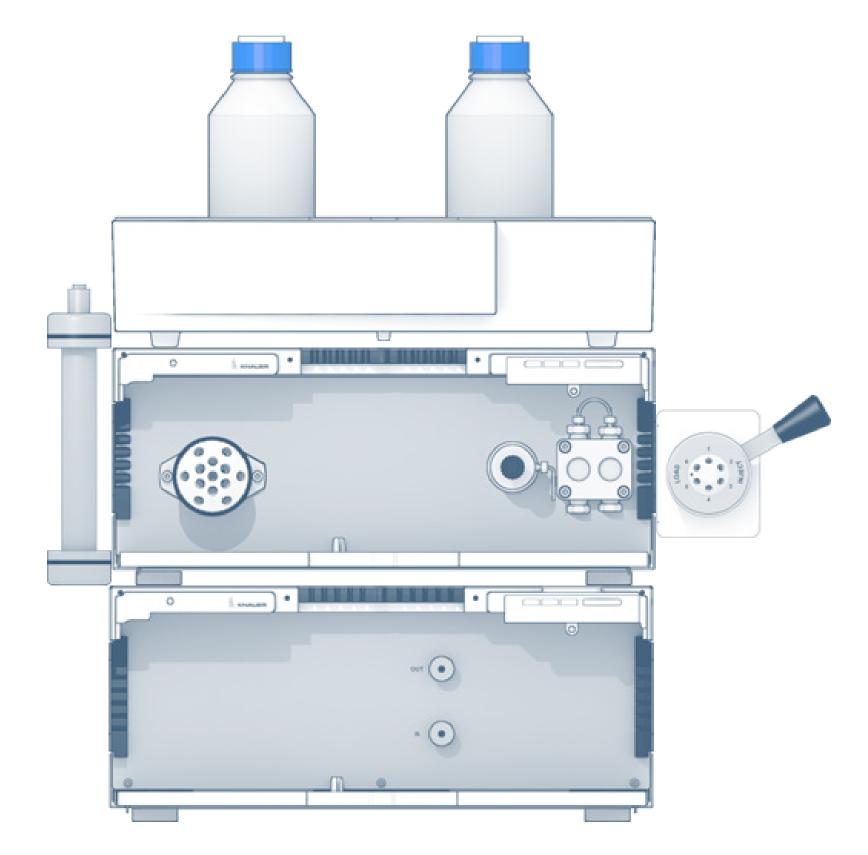
Eluent A	H_2O_{dd}		
Gradient	isocratic, 100 % A		
Flow rate	4 mL/min	System pressure	5.5 bar
Column temperature	60 °C	Run time	16 min
Injection volume	0.5 mL, 1 mL, 2 mL	Injection mode	Full loop
Detection wavelength	RI	Data rate	5 Hz
		Time constant	0.02 sec

Tab. A2 Method parameters (fraction analysis)

Eluent A	H ₂ O _{dd}		
Gradient	isocratic, 100 % A		
Flow rate	0.5 mL/min	System pressure	24 bar
Column temperature	75 °C	Run time	30 min
Injection volume	20 µL	Injection mode	Full loop
Detection wavelength	RI	Data rate	5 Hz
		Time constant	0.05 sec

Tab. A3 System configuration & data (Dedicated sugar purification system)

Instrument	Description	Article No.
Detector	AZURA RID 2.1L	ADD31
Injection	Manual 6-port/3-channel injection valve	A1357
Assistant	AZURA ASM 2.1L: Left: 12 port Multiposition valve as fractionation valve, 8" Middle: - Right: AZURA P 4.1S, 50 mL SSt	AYFALXBD
Heating	Customized heating sleeve, 150 x 20 mm Temperature Control for KNAUER Column Heating Sleeve	A57026 A57024
Column	Eurokat Ca 150 x 20 mm	15PX360EKX
Software	ClarityChrom® Prep 6.1.0	A1685-9



System configuration & data (Analytical system) see Application note VFD0151