

# Group separation with Sepapure Desalting on AZURA® Bio purification system

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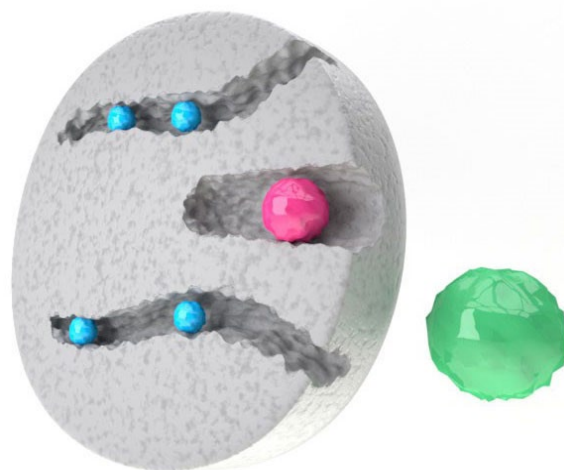
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## SUMMARY

Size exclusion chromatography (SEC) is a popular FPLC technique used for protein purification. Molecules are separated according to their size. Depending on the aim of the purification high resolution fractionation or group separation is used. This application describes group separation with Sepapure Desalting and shows examples for the separation of proteins from dyes and for protein desalting.

## INTRODUCTION

Size exclusion chromatography (SEC) separates molecules according to their different molecular size. In comparison to other chromatography methods, in SEC the sample does not interact with the column matrix. The pore size of the SEC matrix allows the distribution of molecules of different sizes over the column bed and results in separation of the sample. Bigger molecules cannot enter the pores and pass through the column eluting first from the column. The smaller the molecules the better they can enter the pores and therefore have a longer way through the column resulting in a later retention time (**Fig 1**). SEC can be used for high resolution fractionation or group separation of molecules. In group separation the sample is separated into two groups: the high- and low-molecular weight fraction. Group separation can be used for protein purification to remove low molecular weight contaminations like dyes or for desalting and buffer exchange. Here, Fluorescein a popular fluorescent derivatization reagent used for labeling of biomolecules was removed after the labeling process by SEC. Another common use for SEC is desalting. Proteins of interest are not retained by the column and elute first. The small salt molecules elute later and are thereby separated from the



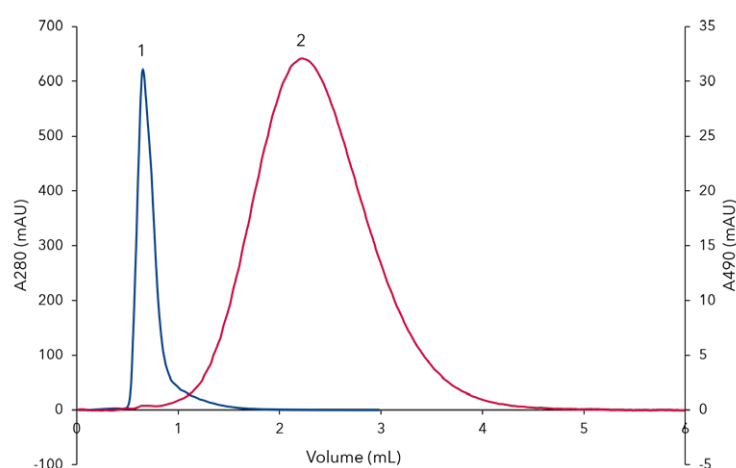
**Fig. 1** Principle of size exclusion chromatography

sample. This mechanism can be used as well for buffer exchange, dyes or for desalting and buffer exchange. Here, Fluorescein a popular fluorescent derivatization reagent used for labeling of biomolecules was removed after the labeling process by SEC. Another common use for SEC is desalting. Proteins of interest are not retained by the column and elute first. The small salt molecules elute later and are thereby separated from the sample. This mechanism can be used as well for buffer exchange.

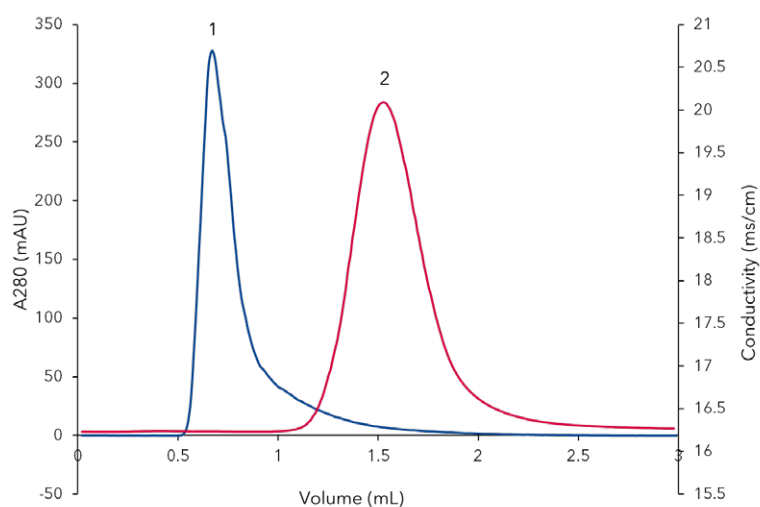
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## RESULTS

In the first method bovine Serumalbumin (BSA) was separated from 5-Carboxyfluorescein (5-FAM) (**Fig 2**). The high molecular weight compound BSA (Peak 1) eluted before the low molecular weight molecule 5-FAM (Peak 2) from the Desalting column. In the second method (**Fig 3**) BSA (Peak 1) was separated from NaCl (Peak 2).



**Fig.2** Separation of BSA and 5-FAM. Peak 1 BSA, Peak 2 5-FAM, red signal UV 280nm, blue signal UV460 nm



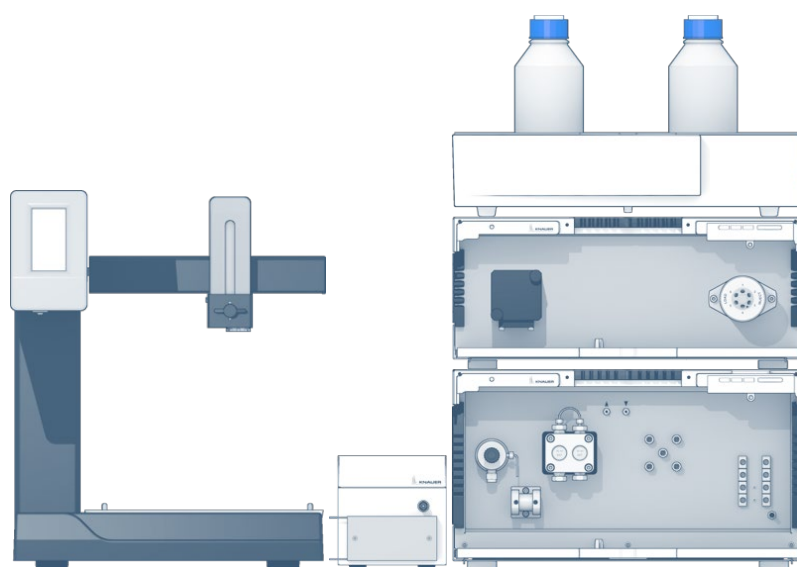
**Fig.3** Desalting of BSA. Peak 1 BSA, Peak 2 NaCl, blue signal UV280 nm, red signal conductivity

## MATERIALS AND METHOD

In this application, an AZURA Bio purification system consisting of AZURA P 6.1L LPG metal-free pump with 10 mL pump head; AZURA ASM 2.1L assistant module with an injection valve and a single wavelength UV detector UVD 2.1S; AZURA CM 2.1S conductivity monitor and Foxy R1 fraction collector was used. 1 mg bovine serum albumin (BSA) and 3.75  $\mu\text{g}$  5-Carboxyfluorescein (5-FAM) was dissolved in PBS. Prior to the run the 1 mL Sepapure Desalting column was equilibrated with PBS. 50  $\mu\text{L}$  of the sample was injected with a flowrate of 1 mL/min. BSA was detected at 280 nm, 5-FAM was detected at 490nm and conductivity signal was recorded to monitor the salt peak.

## CONCLUSION

Sepapure Desalting can be used for the separation of small from large molecules. BSA was separated from a fluorescent dye. Additionally, the buffer was changed by a desalting step. These two examples illustrated the principle of group separation by SEC.



## ADDITIONAL MATERIALS AND METHODS

**Tab. A1** Method parameters

Buffer A	Washing buffer: PBS (phosphate buffered saline)		
Gradient	isocratic		
Flow rate	1 mL/min	System pressure	<3 bar
Column temperature	RT	Run time	6 min
Injection volume	Each 50 µL	Injection mode	-
Detection wavelength	280 nm 490 nm	Data rate	2 Hz

**Tab. A2** System configuration

Instrument	Description	Article No.
Pump	AZURA P6.1L LPG, 10 ml PEEK	<a href="#">APH69EB</a>
Assistant	AZURA ASM 2.1L Right: UVD2.1S Middle: - Left: V2.1S 6 Port/ 2Position	<a href="#">AYCALXEC</a>
Flow cell	3 mm semiprep, 2 µL biocompatible	<a href="#">A4045</a>
Conductivity monitor	CM 2.1S	<a href="#">ADG30</a>
Flow cell	Preparative up to 100 mL	<a href="#">A4157</a>
Column 2	Sepapure Desalting 5 mL	<a href="#">020X460SPZ</a>
Fraction collector	FoxyR1	<a href="#">A59100</a>
Software	PurityChrom, standard licence	<a href="#">A2650</a>

## RELATED KNAUER APPLICATIONS

**VBS0070** - Ion Exchange Chromatography with AZURA® Bio purification system

**VBS0071** - Comparison of two column sets for antibody purification in an automated two step purification process

**VBS0072** - Separation of proteins with cation exchange chromatography on Sepapure SP and CM

**VBS0073** - Separation of proteins with anion exchange chromatography on Sepapure Q and DEAE

**VBS0074** - Comparison of Ion Exchange columns